

THERAPEUTIC ULTRASOUND ASSOCIATED WITH STATIC STRETCHING: EVALUATION OF RATS' SOLEUS MUSCLE SUBMITTED TO REMOBILIZATION PROTOCOL.

⁽¹⁾Alberito Rodrigo de Carvalho, ⁽¹⁾Daniela Martins Cunha, ⁽¹⁾Cassiane Merigo Nascimento, ⁽¹⁾Deisi Ferrari, ⁽¹⁾Gladson Ricardo Flor Bertolini, ⁽²⁾Lucinéia de Fátima Chasko Ribeiro.

⁽¹⁾Laboratory for Study of Injury and Physiotherapy Resources / UNIOESTE – Cascavel-PR, Brazil. ⁽²⁾Medical and Pharmaceutical Sciences Center / UNIOESTE - Cascavel-PR, Brazil. Email: albertorodriigo@gmail.com

ABSTRACT: Despite the deleterious effects, immobilization, is still often used in the treatment of musculoskeletal disorders, even with the occurrence of muscular atrophy, its reversal is a major challenge to rehabilitation. The aim of this study was evaluate the tropism of rats soleus muscle submitted to remobilization with static stretching, preceded by ultrasound, thermal and non thermal. We used 28 rats divided into four groups: G1 – immobilized and remobilized with static stretching, G2 – remobilized with stretching preceded by ultrasound 1.0 W/cm²; G3 – ultrasound at 0.5 W/cm² , G4 – ultrasound with 0.2 W/cm². All animals were immobilized in plantarflexion, producing shortening of the right soleus muscle for 15 days. For the groups subjected to remobilization with ultrasound doses were used according to the group, for 3 minutes, for 10 days with an interval of 2 days after the 5th treatment. After treatment with ultrasound (or not for the G1), the animals were subjected to 3 sets of 30, with 30s interval between them, of static stretching of the soleus. The soleus were dissected, weighed and processed for preparation of histological slides in cross section, and evaluated the smallest diameter of 100 fibers per muscle. There was significant reduction in weight between left and right muscles in all groups, for diameters G3 showed no difference. Conclusion: stretching with or without ultrasound, was unable to reverse the deleterious effects of immobilization on muscle weight, but in the mean dose there was protective effect on the diameter of the fibers.

Keywords: skeletal muscle, muscular atrophy, muscle stretching exercises, ultrasonic therapy, physical therapy modalities.

INTRODUCTION

Skeletal muscles respond to different stimuli, including immobilization. The inactivity causes significant muscle remodeling, including loss of myofibrillar proteins and changes in metabolic activity^[1]. Despite the deleterious effects, immobilization is still often used in the treatment of musculoskeletal disorders, even with the occurrence of muscle cells atrophy and loss of extensibility. The occurrence of muscular atrophy during immobilization and its recovery, presenting a major challenge to rehabilitation^[1].

The therapeutic ultrasound possesses physical, biophysical and therapeutic actions, being target of investigations because of its important roles in metabolic processes and also to repair cell damage. However, some effects are still controversial in the literature regarding the therapeutic use of the modality^[2]. There is still a gap regarding the effects of therapeutic ultrasound on muscle mass in the muscles after a period of immobilization. Therefore, the purpose of this study was to evaluate the soleus muscle mass of rats subjected to remobilization protocol with static stretching, preceded by therapeutic ultrasound, thermal and non thermal.

METHODS

Experimental Groups: We used 28 male albino Wistar rats, with 10 ± 2 weeks old. The study was conducted according to international standards of ethics in animal experiments. The animals were grouped and kept in plastic cages in controlled conditions, with light / dark cycle of 12 hours, temperature 23 ° C ± 2 ° C, with access to water and food *ad libitum*.

The animals were randomly divided into four groups:

• G1 (n=7): animals in this group had immobilized his right ankle in maximum plantar flexion for 15 consecutive days, in order to maintain the soleus muscle in shortened position. After the immobilization period, the animals were subjected to passive stretch of soleus muscle daily for 10 days, with two rest days between the 5th and 6th

therapies; •G2 (n=7): this group was also subjected to immobilization. After this period, the animals were subjected for more two weeks of treatment with therapeutic ultrasound 1 MHz, 1.0 W/cm². Then we performed the static stretching protocol. The procedures were performed similarly to G1; • G3 (n=7): this group was subjected to immobilization and remobilization, similar to previous groups. The intervention was similar to G2, but with a dose of 0.5 W/cm²; • G4 (n=7): this group was subjected to immobilization and remobilization, similar to previous groups. The intervention was similar to G2 and G3, but with a therapeutic dose of 0.2 W/cm².

Immobilization protocol: To conduct the study we used as immobilization apparatus, which aims to shorten the soleus muscle, for this, the tibio-tarsal joint was immobilized in maximal plantar flexion. The animals were observed daily during 15 days of detention in order to repair possible damage to the apparatus. After removal of immobilization, rats were weighed and submitted to the trichotomy of the right posterior region of the triceps surae (soleus).

Ultrasound therapeutic protocol: To perform the therapy with ultrasound device was used Sonopuls Ibramed[®], which had valid calibration certificate during the research period. The frequency used was 1.0 MHz, ERA, 1 cm², and a dose of 1.0 W/cm², 0.5 W/cm² and 0.2 W/cm², respectively, at G2, G3 and G4, for 3 minutes on the region of the right hind limb soleus for 10 therapies, with an interval of two after the 5th treatment. To implement it we used a PVC retainer to immobilize the animal.

Stretching protocol: Subsequent to treatment with ultrasound, static stretch was initiated. To perform the technique of stretching in soleus muscle, the tibio-tarsal joint was maintained manually in maximum dorsiflexion during the entire period of stretching, which consisted of three sets of 30 seconds with a rest interval of 30 seconds between sets. During the stretch animals were kept on PVC retainer.

Histological analysis: At the end of the period of remobilization, all animals were euthanized by guillotine decapitation. Soon after, the right and left soleus muscles were isolated, cleaned and weighed on an analytical balance (Shimadzu®). Were positioned on a Styrofoam mold, fixed with needles on their edges, and remained in formalin 10% until preparation of histological sections, in cross-section of 10 µm, after embedding in paraffin. Then were mounted on the histological sections and subsequently stained with hematoxylin and eosin (HE).

The slides were observed in common light optical microscope (Olympus®), digital camera (EDC-s) attached and a 10x objective lens to perform the digitization of images of muscle fibers transverse sections. Then the images were analyzed using Image-Pro Plus® 3.0, as the smaller diameter of 100 fibers per muscle.

Data analysis: Data were evaluated by comparing the results obtained in the left soleus (intact) and right (subject to immobilization), among the animals of the same experimental group using the Student *t* test, and comparison between groups was performed using one-way ANOVA, with pos hoc of Tukey, and considered significant $p < 0.05$.

RESULTS AND DISCUSSION

In comparing the left soleus muscles weight with the rights, it was possible to observe differences in all groups; when comparing between the groups there was significant difference of the G2 right with the G4 right (fig . 1).

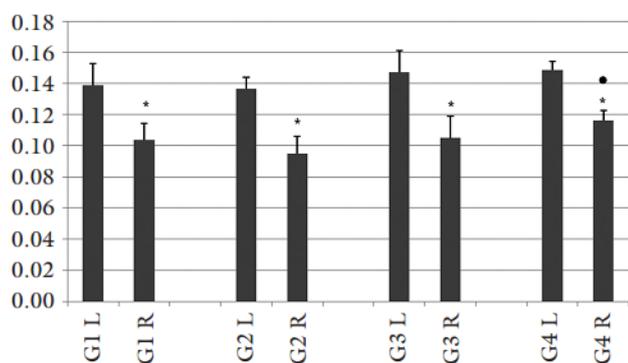


Figure 1 – Graphic representation of means and standard deviations of different groups with respect to the left soleus muscles weight (L) and right (R), according to the different groups. * Statistically significant difference when compared with the contra-lateral. • Significant difference when comparing with G2R.

Comparing the diameters, it was observed that only G3, no significant difference when comparing with the right side with the left. When comparing between groups, to the right side there was no significant difference in any time, but the left side there was a significant difference between G1 and G2 (Fig. 2).

According to Goldspink^[3], the muscle disuse leads to atrophy, because the immobilization of a muscle in shortened position leads to a smaller muscle, since it is metabolically expensive for the organism to maintain a larger muscle than is physiologically necessary. To minimize these effects, uses up resources such as stretch and contractile activity, either by active contraction or

electrical stimulation. However, the present study, we could see signs of muscle atrophy, decreased muscle mass, for the four study groups, indicating that the static stretching alone was not effective in restoring muscle mass, similar to the contralateral side.

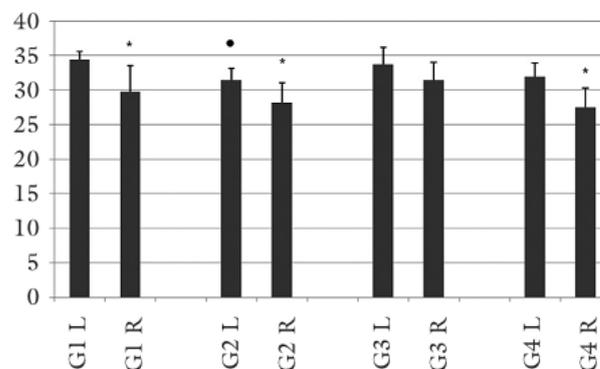


Figure 2 - Graphic representation of means and standard deviations of different groups with respect to the muscle fibers diameters of left soleus muscles (L) and right (R), according to the different groups. * Statistically significant difference when compared with the contra-lateral. • significant difference when comparing with G1L.

It is likely that the mechanical (not thermal) provide benefits in the repair of skeletal muscle^[4], as producing changes in membrane permeability and stimulation of transport by stimulating the proliferation of satellite cells, with formation of new fibers or can assist in the repair of a focal lesion in the early stages of regeneration^[5]. Thus, it is believed that in our results, the ultrasound therapeutic in threshold dose for thermal effects^[2], may have produced catalytic effects on muscle recovery, when combined with stretching.

CONCLUSION

In this study, just the stretching was not enough to reverse the deleterious effects of immobilization, as well as its association with ultrasound in high or low doses, however, using a medium dose, we observed a small protective effect on the diameter soleus muscle fibers.

REFERENCE

- 1 Stevens AE, et al. Muscle adaptations with immobilization and rehabilitation after ankle fracture. *Medicine and Science in Sports Exercise*, **36**:1695-1701, 2004.
- 2 Johns LD. Nonthermal effects of therapeutic ultrasound: the frequency resonance hypothesis. *Journal of Athletic Training*, **37**:293-299, 2002.
- 3 Goldspink DF. The influence of immobilization and stretch on protein turnover of rat skeletal muscle. *Journal of Physiology*, **264**:267-282, 1977.
- 4 Markert CD, et al. Nonthermal Ultrasound and Exercise in Skeletal Muscle Regeneration. *Archives of Physical Medicine and Rehabilitation*, **86**, 2005.
- 5 Karnes JL, et al. Continuous therapeutic ultrasound accelerates repair of contraction-induced skeletal muscle damage in rats. *Archives of Physical Medicine and Rehabilitation*, **83**:1-4, 2002.