INTRODUCTION

Skeletal muscles respond to different stimuli, including immobilization. The inactivity causes significant muscle remodeling, including loss of myofibrillar proteins and changes in metabolic activity[1]. Despite the deleterious effects, immobilization is still often used in the treatment of musculoskeletal disorders, even with the occurrence of muscular atrophy and protein loss. The occurrence of muscular atrophy during immobilization and its recovery, presenting a major challenge to rehabilitation[1].

The therapeutic ultrasound possesses physical, biophysical and therapeutic actions, being target of investigations because of its important roles in metabolic processes and also to repair cell damage. However, some effects are still controversial in the literature regarding the therapeutic use of the modality[2]. There is still a gap regarding the effects of therapeutic ultrasound on muscle mass in the muscles after a period of immobilization. Therefore, the purpose of this study was to evaluate the soleus muscle mass of rats subjected to immobilization protocol with static stretching, preceded by therapeutic ultrasound, thermal and non thermal.

METHODS

Experimental Groups: We used 28 male albino Wistar rats, with 10 ± 2 weeks old. The study was conducted according to international standards of ethics in animal experiments. The animals were grouped and kept in plastic cages in controlled conditions, with light / dark cycle of 12 hours, temperature 23 º C ± 2 º C, with access to water and food ad libitum.

The animals were randomly divided into four groups:

• G1 (n=7): animals in this group had immobilized his right ankle in maximum plantar flexion for 15 consecutive days, in order to maintain the soleus muscle in shortened position. After the immobilization period, the animals were subjected to passive stretch of soleus muscle daily for 10 days, with two rest days between the 5th and 6th therapies; • G2 (n=7): this group was also subjected to immobilization. After this period, the animals were subjected for more two weeks of treatment with therapeutic ultrasound 1 MHz, 1.0 W/cm². Then we performed the static stretching protocol. The procedures were performed similarly to G1; • G3 (n=7): this group was subjected to immobilization and remobilization, similar to previous groups. The intervention was similar to G2, but with a dose of 0.5 W/cm²; • G4 (n=7): this group was subjected to immobilization and remobilization, similar to previous groups. The intervention was similar to G2 and G3, but with a therapeutic dose of 0.2 W/cm².

Immobilization protocol: To conduct the study we used as immobilization apparatus, which aims to shorten the soleus muscle. For this, the tibio-tarsal joint was immobilized in maximal plantar flexion. The animals were observed daily during 15 days of detention in order to repair possible damage to the apparatus. After removal of immobilization, rats were weighed and submitted to the trichotomy of the right posterior region of the triceps surae (soleus).

Ultrasound therapeutic protocol: To perform the therapy with ultrasound device was used Sonopuls Ibramed®, which had valid certification during the research period. The frequency used was 1.0 MHz, ERA, 1 cm², and a dose of 1.0 W/cm², 0.5 W/cm² and 0.2 W/cm², respectively, at G2, G3 and G4, for 3 minutes on the region of the right hind limb soleus for 10 therapies, with an interval of two after the 5th treatment. To implement it we used a PVC retainer to immobilize the animal.

Stretching protocol: Subsequent to treatment with ultrasound, static stretch was initiated. To perform the technique of stretching in soleus muscle, the tibio-tarsal joint was maintained manually in maximum dorsiflexion during the entire period of stretching, which consisted of three sets of 30 seconds with a rest interval of 30 seconds between sets. During the stretch animals were kept on PVC retainer.
Histological analysis: At the end of the period of remobilization, all animals were euthanized by guillotine decapitation. Soon after, the right and left soleus muscles were isolated, cleaned and weighed on an analytical balance (Shimadzu®). Were positioned on a Styrofoam mold, fixed with needles on their edges, and remained in formalin 10% until preparation of histological sections, in cross-section of 10 μm, after embedding in paraffin. Then were mounted on the histological sections and subsequently stained with hematoxylin and eosin (HE). The slides were observed in common light optical microscope (Olympus®), digital camera (EDC-s) attached and a 10x objective lens to perform the digitization of images of muscle fibers transverse sections. Then the images were analyzed using Image-Pro Plus® 3.0, as the smaller diameter of 100 fibers per muscle.

Data analysis: Data were evaluated by comparing the results obtained in the left soleus (intact) and right (subject to immobilization), among the animals of the same experimental group using the Student t test, and comparison between groups was performed using one-way ANOVA, with pos hoc of Tukey, and considered significant p < 0.05.

RESULTS AND DISCUSSION

In comparing the left soleus muscles weight with the rights, it was possible to observe differences in all groups; when comparing between the groups there was significant difference of the G2 right with the G4 rithth (fig. 1).

Figure 1 – Graphic representation of means and standard deviations of different groups with respect to the left soleus muscles weight (L) and right (R), according to the different groups. * Statistically significant difference when compared with the contra-lateral. ● Significant difference when comparing with G2R.

Comparing the diameters, it was observed that only G3, no significant difference when comparing with the right side with the left. When comparing between groups, to the right side there was no significant difference in any time, but the left side there was a significant difference between G1 and G2 (Fig. 2).

According to Goldspink[3], the muscle disuse leads to atrophy, because the immobilization of a muscle in shortened position leads to a smaller muscle, since it is metabolically expensive for the organism to maintain a larger muscle than is physiologically necessary. To minimize these effects, uses up resources such as stretch and contractile activity, either by active contraction or electrical stimulation. However, the present study, we could see signs of muscle atrophy, decreased muscle mass, for the four study groups, indicating that the static stretching alone was not effective in restoring muscle mass, similar to the contralateral side.

CONCLUSION

It is likely that the mechanical (not thermal) provide benefits in the repair of skeletal muscle[4], as producing changes in membrane permeability and stimulation of transport by stimulating the proliferation of satellite cells, with formation of new fibers or can assist in the repair of a focal lesion in the early stages of regeneration[8]. Thus, it is believed that in our results, the ultrasound therapeutic in threshold dose for thermal effects[2], may have produced catalytic effects on muscle recovery, when combined with stretching.

REFERENCE